

## **ISOLATION AND IDENTIFICATION OF HEAVY METAL RESISTANT BACTERIA BASED ON 16S rRNA SEQUENCING FROM BARITE MINES OF KADAPA DISTRICT ANDHRA PRADESH.**

**J Patricia Raj kumari and Dr M Nagalakshmi Devamma\***

Department of virology, S. V. University, Andhra Pradesh, Tirupati -517502, India.

\*Assistant Professor, Dept of Botany, S. V. University, Tirupati -517502.

Phone No: 91-9676318138, 91-7569242750

### **ABSTRACT**

This present study focuses on isolation and screening of a indigenous heavy metal resistant bacteria strain from Agricultural and Mining soils N:14°03'07.4" E 79.18.01.0" ,N : 14.01 32.5" E 79 ° 19'12.7"of Barites mines Mangampet YSR Kadapa District Andhra Pradesh, Individual colonies of bacteria which varied in shape and colour were picked up and purified by streaking on Nutrient agar, four bacteria were screened for heavy metal resistance succeeding isolation from respective soils they were exposed to different concentrations of Chromium, Lead, Copper, Nickel, Zinc from 0 to 8000mg/l. conclusively a strain outstood in tolerating 8000mg/l of Lead acetate besides growing in other metal solutions for over 300mg/l,  $K_2Cr_2O_7$  ,200mg/l  $NiCl_2$ ,  $CuSO_4$  300mg/l 300mg/l  $ZnCl_2$ . Hence the strain which could tolerate 8000 mg/l of lead acetate felt the need of identification and was identified based on 16S rRNA sequencing as *klebsiella pneumoniae* with Accession number KJ 194599.

**Keywords:** *Klebsiella pneumoniae*, Heavy metal resistant, Barites mining area, YSR Kadapa District.

**Corresponding Author** \*devi.bot@gmail.com

### **INTRODUCTION**

Heavy metals is a general collective term to the group of metals and metalloids with atomic density greater than 4000 kg m<sup>-3</sup> or 5 times more than water however some of them act as essential micro nutrients for living beings but when it exceeds in concentrations they can be harmful. In the environment the heavy metals continue to exist for a longer period and are stable than organic contaminants and are non- biodegradable The environmental pollution by heavy metals arrives from anthropogenic sources such as smelters, mining, power stations and the application of pesticides containing metal fertilizer and sewage sludge and the reckless disposal of wastes by various industries they can become mobile in soils depending on soil pH and their speciation So a fraction of the total mass can leach to aquifer or can become bio available to living organisms heavy metals can stagnate in biological systems and ultimately be introduced into food web via different mechanisms them.[15-19] Microbes which are continuously exposed to heavy metal stress have adapting mechanisms to the metal

contaminants, have a multiple ways to carry on with high concentrations of heavy metals and often are specific to one or a few metals [20-24] Indigenous organisms have adapted to novel environments but have also thrived under them.[4-5] They possess a variety of mechanisms to deal with higher concentrations of heavy metals because they have developed mechanisms to tolerate heavy metals through efflux, complexation, or reduction of metal ions or to use them as terminal electron acceptors in Anaerobic respiration.[7] Microorganism able to survive well in higher concentrations of heavy metals are of great interest as bioremediation agents because they can achieve different transformation and immobilization processes. Specifically they bioaccumulate based on the incorporation of metals inside the living biomass or biosorption in which metal ions are adsorbed at the cellular surface by different mechanisms[6]. Metal accumulative bioprocess generally falls into one of two categories, absorptive(passive) uptake by nonliving, non growing biomass or biomass products and bioaccumulation by living cell [31-36] and the complex structure of microorganisms implies that there are many ways for the metal to be taken up by the microbial cell according to the dependence on the cell's metabolism[1]. Many microorganisms have developed chromosomally or extra chromosomally controlled detoxification mechanisms to overcome the detrimental effects of heavy metals[3]. The detoxifying ability of these resistant microorganisms can be applied for bioremediation of heavy metals in wastewater. Effluents having heavy metals can be treated with these microorganisms by the processes like bio-sorption, bioaccumulation and bio precipitation. This study aimed to isolate and identify heavy metal resistant bacteria tolerating higher concentrations of heavy metal from Barite Agricultural and mining area situated at Mangampet YSR kadapa district Andhra Pradesh and its application like above.

## **ISOLATION OF BACTERIA FROM SAMPLES**

### **Study area and sampling**

Soil samples were collected from Barite Mining and Agricultural area Mangampet Y.S.R District. Samples were collected from the depth of approximately 15 cm after which they were placed in sterilized polyethylene bags using a sterilized spatula. All samples were stored at 4 °C until analysis.[ 24]

## **CHEMICALS AND MEDIA**

Stock solutions of heavy metal (1000 mg L<sup>-1</sup>) were prepared in distilled water. The elemental salts employed were lead acetate Pb(C<sub>2</sub>H<sub>3</sub>O<sub>2</sub>)<sub>2</sub>, Potassium dichromate (K<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub>), Nickel chloride (NiCl<sub>2</sub>), Zinc chloride (ZnCl<sub>2</sub>), copper sulphate (CuSO<sub>4</sub>), (from DDH, Mumbai India). Nutrient agar was used for isolation of bacteria. [35, 42]

## **ISOLATION OF BACTERIA**

Soil samples were passed through a sieve (2 mm) to remove large pieces of debris and vegetation. The bacteria were originally isolated by plating dilutions of soils in distilled water on Nutrient agar and then incubated at 30 °C for 48 h. Individual colonies of bacteria which varied in shape and colour were picked up and purified by streaking on nutrient agar.

The bacterial isolates were maintained on nutrient agar at 4°C and recultured every four weeks. All isolated, selected bacteria were purified and investigated for further work. [45]

## SCREENING OF HEAVY METAL RESISTANT BACTERIA

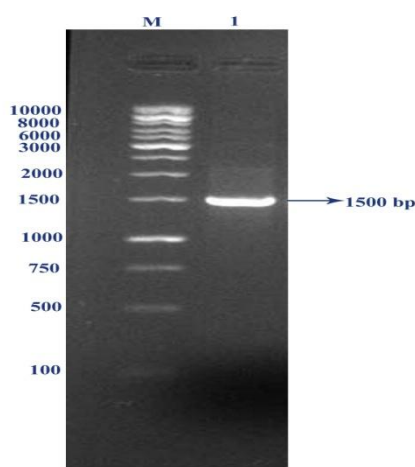
The four isolated strains were screened for resistance using the Agar diffusion method. Agar plates containing 20 ml nutrient agar medium supplemented with different concentrations from (0 to 8000mg/l) lead acetate  $Pb(C_2H_3O_2)_2$ , potassium dichromate ( $K_2Cr_2O_7$ ) and Nickel chloride ( $NiCl_2$ ), zinc chloride ( $ZnCl_2$ ), copper sulphate ( $CuSO_4$ ) were inoculated with the purified bacteria and incubated at 30 °C for 48h[35]. An Isolate showed high resistance to lead which is at concentration of 8000mg/l besides 300mg/l  $K_2Cr_2O_7$ , 200mg/l  $NiCl_2$ ,  $CuSO_4$  300mg/l, 300mg/l  $ZnCl_2$ . and was therefore identified based on its 16S rRNA gene of selected isolate.[45,42]

Resistance of bacterial isolates to different heavy metals and concentrations

Elements	Conc./mg/l	X2	X3	X4
Lead	50	+	+	+
	100	-	-	+
	200	-	-	+
	300	-	-	+
	40 to 8000mg/l	-	-	+
Chromium	50	-	-	+
	100	-	-	+
	200	-	-	+
	300	-	-	+
	400 to 8000mg/l	-	-	-
Zinc	50	+	+	+
	100	+	+	+
	200	-	-	+
	300	-	-	+
	400 to 8000mg/l	-	-	-
Nickel	50	-	-	+
	100	-	-	+
	200	-	-	+
	300	-	-	-
	400 to 8000mg/l	-	-	-
Copper	50	-	-	+
	100	+	-	+
	200	+	+	+
	300	-	-	+
	400 to 8000mg/l	-	-	-

## ANALYSIS OF 16S rRNA GENES WAS CONDUCTED AS

The method described by Ausubel et al(1994)[2] was slightly modified and used for Genomic DNA isolation. PCR was performed for the amplification of 16S region. Amplification was performed using the following Amplification conditions: Amplification Step 1: 94°C for 5 min [Initial denaturation], : 94°C for 1 min [Denaturation] Step 3: 55°C for 1 min (annealing) 40 cycles, Step 4: 72°C for 1 min (Elongation) Step 5: 72°C for 10 min (final extension). DNA oligomers used in this experiment were Forward 16sF: 5'AGAGTTGATCCTGGCTCAG-3' Reverse 16sR: 5'-CAAGGCATCCACCGT3'. The forward primer was complementary to the upstream of 16S rDNA. The PCR products were visualized on an UV illuminator (UV Tech, France). The images of gels were further analysed by a Image J computer programme.



## SEQUENCING

The sequencing reactions were performed in a SEQ4×4 Personal Sequencing System (MWG Biotech, Bangalore). The sequences obtained were then evaluated in GenBank using BLAST programme and accession numbers were obtained.

## RESULTS AND DISCUSSION

In this study different heavy metals which are known to be toxic viz, chromium, lead, were used. Presence of heavy metal tolerant bacterium in a particular environment may be an indication that such area is affected by heavy metals, Such an area may endorse heavy metal resistant organisms [11]. Bacteria are primarily the first ones to be affected by discharges of heavy metals into the environment resulting in an increase of heavy metal resistant bacteria in these environments [9, 20, 21, 10]. Isolation of bacteria from metal polluted environment would represent an appropriate practice to select metal resistant strains that could be used for heavy metal removal and bioremediation purposes [12]. Microorganisms undergo selection pressures in the presence of toxic compounds and develop resistance [13]. In the current study in Barite mining area Bacteria varied in colour and shape were picked for screening of heavy metals from isolated bacteria from the soils of respective places mentioned above. The purified isolates were tested for their resistance to heavy metals like Nickel, Chromium, Zinc, Lead, Copper, using the Agar diffusion method on Nutrient agar medium. [35] The degree of heavy metals resistance to the highest concentration in the nutrient agar media was evaluated.

based on the ability of the isolated bacteria to grow on the subsequent higher concentrations. Among the bacterial isolates *Klebsiella pneumoniae* (KJ 194599) tolerated  $K_2Cr_2O_7$  300mg/l,  $NiCl_2$  200mg/l,  $CuSO_4$  300mg/l,  $ZnCl_2$  300mg/l, lead at the maximum 8000 mg/L. The difference in the toxicity toward the bacterial isolates could be explained by the conditions of bacterial isolation and the nature and physiological characteristics of each bacterial isolate [35]. In high concentrations, heavy metal ions react to form toxic compounds in cells, [22]. It must first enter the cell. Because some heavy metals are necessary for enzymatic functions and bacterial growth, uptake mechanisms exist that allow for the entrance of metal ions into the cell. There are two general uptake systems: one is quick and unspecific driven by a chemiosmotic gradient across the cell membrane and thus requiring no ATP, and the other is slower and more substrate-specific, driven by energy from ATP hydrolysis. While the first mechanism is more energy efficient, it results in an influx of a wider variety of heavy metals, and when these metals are present in high concentrations, they are more likely to have toxic effects once inside the cell [23]. To survive under metal-stressed conditions, bacteria have evolved several types of mechanisms to tolerate the uptake of heavy metal ions. These mechanisms include the efflux of metal ions outside the cell, accumulation and complexation of the metal ions inside the cell, and reduction of the heavy metal ions to a less toxic state [22], whereas in bacteria, various peptides consisting of metal-binding amino acids (mainly histidine and cysteine residues) have been studied for enhanced heavy metal accumulation by bacteria. Various peptides consisting of metal-binding amino acids, mainly histidine and cysteine residues, have been studied for enhanced heavy metal accumulation by bacteria. Novel metal-binding peptides might offer a higher affinity, higher metal-binding capacity, and/or specificity and selectivity for a target metal ion than known metal-binding proteins [33]. The most important of these groups are Carbonyl (ketone), Carboxyl, Sulfhydryl (thiol), Sulfonate, Thioether, Amine, Secondary amine, Amide, Imine, Imidazole, Phosphonate, Phosphodiester [39]. Duxbury, (1986) have opined that the tolerance to heavy metals is seemed to be remarkable in Gram Negative bacteria only. [44]. Thus the organisms isolated and identified from a heavy metal polluted environment developed the mechanisms to survive a highly toxic environment. This study thus highlights the presence of bacteria which survives in a highly heavy metal polluted environment. These isolates are of interest for molecular characterization of mechanisms for resistance to multiple metals and hold promise for bioremediation of toxic heavy metals, including in environments that are contaminated by several metals.

## REFERENCES

1. Ahalya N, Ramachandra TV, Kanamadi RD, "Biosorption of heavy metals. Res." J. Chem. Environ" (2003), 7: 71-78.
2. Asubel, F. H, R. Brent, R. E. Kingston, D. D. Moore, J. G. Seidman, J. A. Smith & K. Struhl. Current protocols in Molecular Biology (1994), John Wiley and Son.
3. Ehrlich, H. L. "Microbes and metals". Applied Microbiology and Biotechnology, (1997), 48: pp 687-692.
4. Haq R, Shakoori AR "Microorganisms resistant to heavy metals and toxic chemicals as indicators of environmental pollution and their use in bioremediation." Folia Biol (Krakow), (2000), 48: 143-147.

5. Roane TM & Pepper IL, "Microbial responses to environmentally toxic cadmium" *Microb Ecol*, (2000), 38: 358–364.
6. Vijayaraghavan, K, Yun, Y. S, "Bacterial bio sorbents and bio sorption." *Biotechnology Advance*, (2008) , 26, 266- 291.
7. Haferburg G, Kothe Metallomics: lessons for metalliferous soil remediation, *Appl. Microbiol. Biotechnology E*, (2010), 87: 1271-1280.
- 8 . Silver S & Phung LT , " Bacterial heavy metal resistance: New surprises." , (1996), *Annu Rev Microbiol*, 50: 753–789.
9. Trevor's, J. "Copper resistance in bacteria *Microbiological Sciences*, (1987)," 4: pp 29-31.
- 10.Nair S. Loka Bharathi, P.A and Chandramohan,D."Effect of Heavy metals on *Bacillus* spp, and *Flavobacterium* spp *Ecotoxicology* , " , (1993), 2pp 220-229.
- 11.Clausen, C.A., "Isolating metal tolerant bacteria capable of removing copper,chromium and arsenic from treated wood" *Waste Management Research* , (2000), 18pp: 264- 268
- 12.Malik,A."Metal bioremediation through growing cells" *Environment International*,(2004), 30: pp 261-278.
- 13.Hideomi,N., IshikawaT., Yasunaga.S., Kondo.I and Mitsuhasi,S."Frequencyof heavymetal resistance in bacteri from inpatients in Japan"(1977), *Nature*. 266: pp 165-167
- 14 .Gabardine JR., Hayes H, Roth D et al., "Contaminants in the Mississippi river. U. S Geological Survey Circular", (1995), Virginia, U.S.A. 1133.
15. Kavamura VN & Esposito E, "Biotechnological strategies applied to the decontamination of soils polluted with heavy metals". *Biotechnology Advances* (2010), 28: 61–69.
16. Lasat MM "Phytoextraction of toxic metals: a review of biological mechanisms", *J. Environ.* (2002), Qual. 31: 109–120.
17. Alloway BJ, "Soil Processes and the Behaviour of Metals". In: Alloway BJ ed. *Heavy Metals in Soils* BJ Blackie & Son Inc. (1990), New York, pp 7-28.
- 18 .Santana L, Castaldi P & Melis P , "Evaluation of the interaction mechanisms between redmuds and heavymetals" . *Journal of Hazardous Materials* (2006), 136, 324-329.
- 19.Giller K E, Witter E, McGrath SP, " Toxicity of heavy metals to microorganisms and microbial process in agricultural soils", *A Review soil Biol Biochem* (1998), 30:1389-1414.
20. Jain, R.K, "Copper resistant microorganisms and their role in the environment" *World Journal of Microbiology and Biotechnology*, (1990), 6: pp 356–365.
21. Silver, S., (Bacterial resistances to toxic metal ions A Review" *Genetics*, (1996)," 179: pp 9-19.



22. Nies, D.H., "Microbial heavy metal resistance." *Appl Microbiol Biotechnology* (1999), 51: 730-750.
23. Nies, D.H., and Silver, S., "Ion efflux systems involved in bacterial metal resistances" *Journal of Industrial Microbiology* (1995).14: 186-199.
24. Laura-Dorina-Dinu ,Laura Anghel, Stefana, JurcoaneJurcoane, " Isolation of heavy metal resistant bacterial strains from the battery manufactured polluted environment Romanian Biotechnological Letters, ", (2011), Vol. 16, No.69(102-106)
25. Silver S & Mistra TK "Plasmid mediated metals resistance. *Annu Rev Microbiol*" (1988), 42: 717-743. 21.
26. Silver S & Phung LT "Bacterial heavy metal resistance": New surprises. *Annu Rev Microbiol* (1996). , 50: 753-789.
27. Mejare M & Bulow L "Metal-binding proteins and peptides in bioremediation and phytoremediation of heavy metals. *Trends Biotechnol*, (2001). 19: 67-73.
28. Nies DH "Efflux-mediated heavy metal resistance in prokaryotes" *FEMS Microbiol Rev*, (2003). 27: 313-339
29. Piddock LJ "Multidrug-resistance efflux pumps-not just for resistance". *Nat Rev Microbiol* (2006)., 4: 629-636
30. R. Munoz, M. T. Alvarez, A. Munoz, E. Terrazas, B. Guieysse, and B.Mattiasson: "Sequential removal of heavy metal ions and organic pollutants using an algal-bacterial consortium". *Chemosphere*, .2006. 63, 903-911
31. Silver, S. Bacterial heavy metal detoxification and resistance systems. In *Biotechnology and Environmental Science: Molecular approaches* (Mongkolsuk, S. *et al.*, eds) (1992, pp. 109-129, Plenum Press.
32. Silver, S. Bacterial heavy metal resistance: New surprises. *Annu. Rev. Microbiol.*50, (1996) 753-789.
33. Wagner, G.J Accumulation of cadmium in crop plants and its consequences to human health.*Adv. Agronomy*. (1993) 51, 173-212.
34. Volesky B. Bio sorption and me. *Water Res* 2007; 41:4017-29.
35. Hassan, S. H. A., Abskharon, R. N. N., Gad El-Rab, S.M. F., Shoreit, A. A. MI Isolation, characterization of heavy metal resistant strain of *Pseudomonas aeruginosa*. Isolated from polluted sites in Assiut city, Egypt. *Journal of Basic Microbiology*. (2008)48: 168-176..
36. Macaskie L and dean ACR, 1989. "Microbial metabolism desolubilisation and deposition of heavy metals metal uptake by immobilization cells and application to the detoxification of liquid wastes", *Adv.Biotechno, proc*, 12:159-172.

37. Aksu Z and Kutsal T, 1990. "A comparative study for bio sorption characteristics of heavy metal ions with *C. vulgaris*. *Environ. technol*, 11: 979-987.
38. Huang C, Huang C and Moreheart AL, 1990. The removal of copper from dilute aqueous solutions by *Saccharomyces cerevisiae*, *Wat. Res.*, 24: 433-439.
39. Volesky B, May H and Holan ZR, 1992, cadmium biosorption by *Saccharomyces Cerevisiae*, *Biotechnol. Bioeng.*, 41: 826-829.
40. Avery SV and Tobin JM, 1993. Mechanism of adsorption of hard and soft metal ions to *Saccharomyces cerevisiae* and influence of hard and soft anions *Appl Environ. Microbiol*, 59: 2851 – 2856.
41. Brady D., Stoll A.D., Starke L and Duncan JR, , " Bioaccumulation of metal cations by *Saccharomyces cerevisiae* *Appl. Microbiol. Biotechnol.* 41:149-154.
42. Virender singh, P K Chauhan, Seema, Ankur Tyagi, Keshav, Thakur, Anu Kumar And Vivek Kumar " Isolation and antibiogram pattern of *E. coli* isolates having heavy metals tolerance" *Jul-Sep. 2010 Vol.1/Issue-3*.
43. Wagner, G.J. (1993) Accumulation of cadmium in crop plants and its consequences to human health. *Adv. Agronomy* 51, 173–212.
44. Duxbury, T. Microbes and heavy metals: An ecological overview. *Microbiology Science*, 1986, 3: 330-333.
45. Velusamy, P., Y. M. Awad, S. A. M. Abd El-Azeem and Y. S. Ok, "Screening of Heavy Metal Resistant Bacteria Isolated from Hydrocarbon Contaminated Soil in Korea" *Journal of Agricultural, Life and Environmental Sciences* Vol.23 No.1 March, 2011